



Research paper

Massively parallel sequencing of 68 insertion/deletion markers identifies novel microhaplotypes for utility in human identity testing



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ABSTRACT

Short tandem repeat (STR) loci are the traditional markers used for kinship, missing persons, and direct comparison human identity testing. These markers hold considerable value due to their highly polymorphic nature, amplicon size, and ability to be multiplexed. However, many STRs are still too large for use in analysis of highly degraded DNA. Small bi-allelic polymorphisms, such as insertions/deletions (INDELs), may be better suited for analyzing compromised samples, and their allele size differences are amenable to analysis by capillary electrophoresis. The INDEL marker allelic states range in size from 2 to 6 base pairs, enabling small amplicon size. In addition, heterozygote balance may be increased by minimizing preferential amplification of the smaller allele, as is more common with STR markers. Multiplexing a large number of INDELs allows for generating panels with high discrimination power. The Nextera™ Rapid Capture Custom Enrichment Kit (Illumina, Inc., San Diego, CA) and massively parallel sequencing (MPS) on the Illumina MiSeq were used to sequence 68 well-characterized INDELs in four major US population groups. In addition, the STR Allele Identification Tool: Razor (STRait Razor) was used in a novel way to analyze INDEL sequences and detect adjacent single nucleotide polymorphisms (SNPs) and other polymorphisms. This application enabled the discovery of unique allelic variants, which increased the discrimination power and decreased the single-locus random match probabilities (RMPs) of 22 of these well-characterized INDELs which can be considered as microhaplotypes. These findings suggest that additional microhaplotypes containing human identification (HID) INDELs may exist elsewhere in the genome.

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1. Introduction

Short bi-allelic insertion/deletion (INDEL) polymorphisms are the second most abundant polymorphism, discovered to date, in humans and have been demonstrated as a useful alternative to traditional short tandem repeat (STR) testing in forensic genetics [1–6]. Due to their small amplicon size, INDELs are more advantageous than STRs for typing compromised DNA samples.

The small difference in allele size potentially minimizes preferential amplification of smaller size alleles of a heterozygote, a more common occurrence with traditional STR testing. INDELs also have relatively low mutation rates and do not generate stutter products during PCR amplification. Lastly, the ease of multiplexing INDELs enables the development of panels with relatively low random match probabilities (RMPs) for human identity (HID) testing [7,8].

Massively parallel sequencing (MPS, also referred to as next generation sequencing (NGS)) is capable of targeting many loci, including those of forensic relevance, across the genomes of multiple samples simultaneously with relatively high sequence coverage [9–15]. With sequencing, it is possible to define INDELs better and potentially identify proximal single nucleotide polymorphisms (SNPs) that can increase the discrimination power of

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currently defined INDELs, i.e., by identifying INDEL-containing microhaplotypes. Herein, the Nextera™ Rapid Capture Custom Enrichment Kit was used to prepare DNA libraries that were sequenced on the Illumina MiSeq to generate sequence data for 68 well-described forensically relevant HID INDELs in four major US population groups. In addition, the STR Allele Identification Tool: Razor (STRait Razor) software [16] was used in a novel way to analyze INDEL sequences and detect adjacent SNPs. This application has enabled the discovery of unique allelic variation, which increases the discrimination power and decreases the single-locus random match probabilities of 22 of the INDELs. The results presented here demonstrate the utility of MPS for typing INDEL flanking regions to increase the discrimination power of current bi-allelic markers for HID testing.

2. Materials and methods

2.1. Samples and DNA extraction

DNA was extracted from whole blood and saliva samples obtained from 190 unrelated individuals following the University of North Texas Health Science Center Institutional Review Board Approval. The sample set represented unrelated individuals of four major U.S. population groups with 49 Caucasians (CAU), 49 African Americans (AFA), 49 Hispanics (HIS), and 43 Asians (ASA). DNA extraction was performed using the Qiagen® QIAamp™ DNA Blood Mini Kit (Qiagen, Hilden, Germany), according to the manufacturer's protocol [17].

2.2. Library preparation and massively parallel sequencing

Libraries were generated using a custom designed Nextera™ Rapid Capture Enrichment panel (Illumina, Inc., San Diego, CA) using the Illumina Design Studio, as described by Zeng, et al. [18] and Warshauer, et al. [19]. Capture probe sequences will be made available upon request. The HID INDELs for this study were selected based on the results described by LaRue, et al. [3] and Pereira, et al. [6]. INDEL rs number, location, flanking region, and probe design are listed in Supplemental Table 1 [20]. 50 ng of genomic DNA were used as input for each library preparation reaction. Each sample library was diluted to 2 nM and paired-end sequencing (12 pooled libraries per run) was performed on the Illumina MiSeq according to the manufacturer's recommended protocol with a read length of 250 bases [21].

2.3. STRait Razor design

A configuration file was created for use with STRait Razor v2.5 (Supplemental Fig. 1 and Supplemental Table 2) [16]. To create the file, locus coordinates for each INDEL were located on the hg19 human reference genome using the Integrative Genomics Viewer (IGV) [22,23]. STRait Razor flanking regions up and downstream of the INDEL motif, and the complementary sequences, were recorded. The average size of the STRait Razor flank, used to mine sequence data for regions of interest, was $24 \text{ bases} \pm 0.10$. The bases between STRait Razor flanks contained the INDEL motif and approximately 50 bases on either end. The STRait Razor flanks

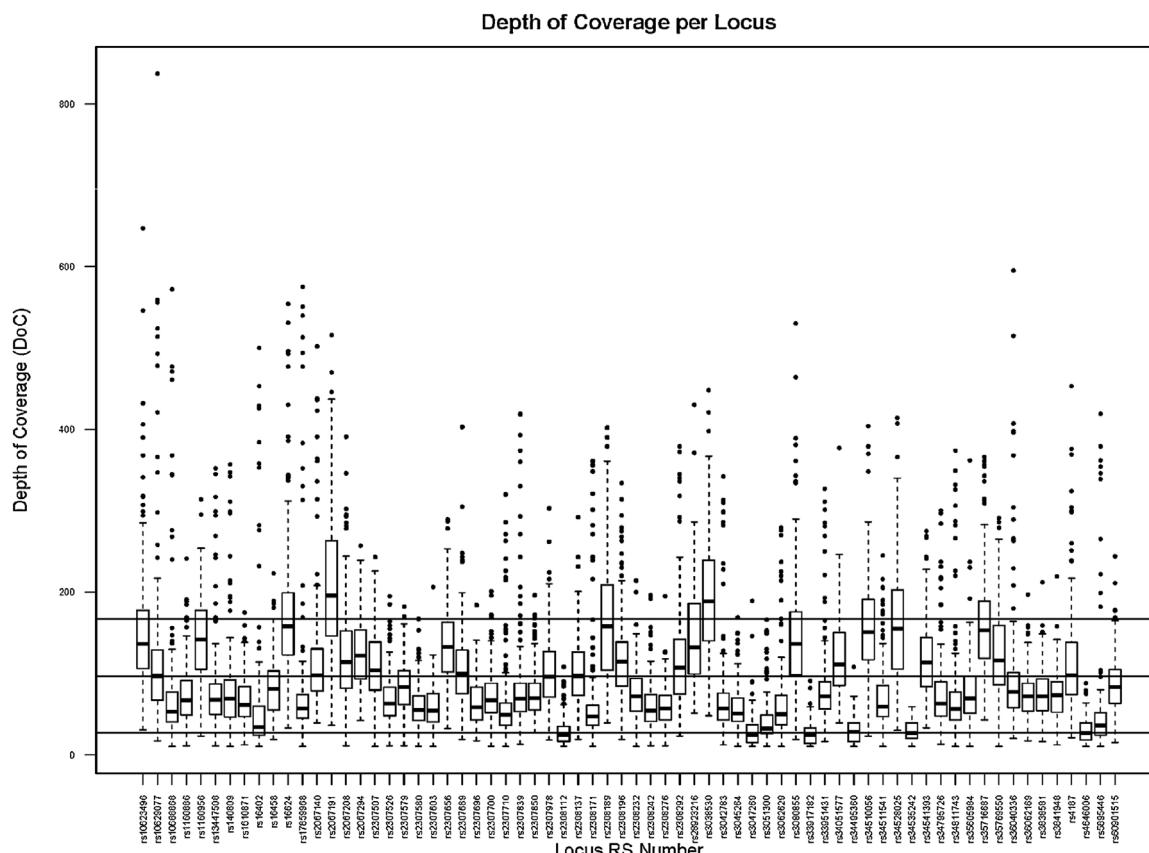


Fig. 1. Depth of coverage (DoC) values for 68 human identity INDELs using the Nextera™ Rapid Capture Enrichment kit and the Illumina MiSeq. Each box plot represents a single locus; the center horizontal line represents the median, the lower and upper boundaries of each box represent the first and third quartiles, respectively, the vertical dashed lines indicate plus/minus three times the interquartile range, and closed circles indicate outliers. The center horizontal line indicates the mean across 68 loci and the top and bottom horizontal lines indicate plus and minus one standard deviation, respectively for all loci combined.

were designed to capture sequence variation in the flanking regions adjacent to the target INDEL (Supplemental Tables 3 and 4) while keeping total target size relatively small. The average length of this region (target INDEL plus approximately 50 bases on either end) was 99 bases \pm 4 and 102 bases \pm 4 for the deletion and insertion alleles, respectively. Lastly, a relatively short sequence between the STRait Razor flanks, but unique relative to the INDEL motif, was recorded; the average length of these sequences was 12 bases \pm 0.15. Analysis of the resulting data was performed using the STRait Razor Sequence Analysis toolkit to assign genotypes to each sample and compile depth of coverage (DoC) and allele coverage ratio (ACR) data. ACRs were calculated by dividing the DoC of one allele by the DoC of the second allele. An ACR of 1.0 was considered perfectly balanced. It should be noted that the hg19 reference genome was used to design the STRait Razor configuration file, however, the sequences within the file are identical to those in the hg38 reference genome.

2.4. Analysis Concordance

Sixty-nine of the samples were analyzed manually by a second reviewer to confirm STRait Razor allele calls. Fastq files were aligned using Burrows-Wheeler Aligner (BWA) and Sequence Alignment/Map Tools (SAMtools) [24–26]. The resulting binary alignment/map (.bam) files were used as input for the Genome Analysis Toolkit (GATK) [27]. The resulting variant call format (.vcf) files were analyzed using an in-house Excel-based workbook. The workbook assigned genotypes and compiled DoC and ACR data for each sample.

2.5. Population statistical analyses

Length-based and sequence-based allele frequencies, observed and expected heterozygosities, and testing for departures from Hardy-Weinberg Equilibrium (HWE) and linkage disequilibrium (LD) assessments were performed using Genetic Data Analysis (GDA) [28]. An in-house Excel-based workbook was used to generate power of discrimination values and single-locus and combined RMPs.

3. Results and discussion

A total of 190 samples were sequenced. One run, containing 11 African American samples and one Asian sample, performed poorly with insufficient sequencing Q scores (between 10 and 20) for all of read 2 and part of read 1. This run was removed from analysis due to poor sequence quality. Ultimately, 178 samples were analyzed, consisting of 48 Caucasians, 38 African Americans, 49 Hispanics, and 43 Asians.

3.1. Locus performance

Analysis of the resulting data was performed using operationally selected DoC and ACR thresholds of 10 x and 0.20, respectively. Mean profile completion was $96.3\% \pm 0.108$, ranging from 44.1% to 100% for the 178 samples. Full HID INDEL profiles were obtained for 70 samples. The average DoC and ACR for 68 HID INDELS was $96.9x \pm 69.9$ and 0.727 ± 0.182 , respectively (Figs. 1 and 2). One locus, rs33917182, fell below one standard deviation from the

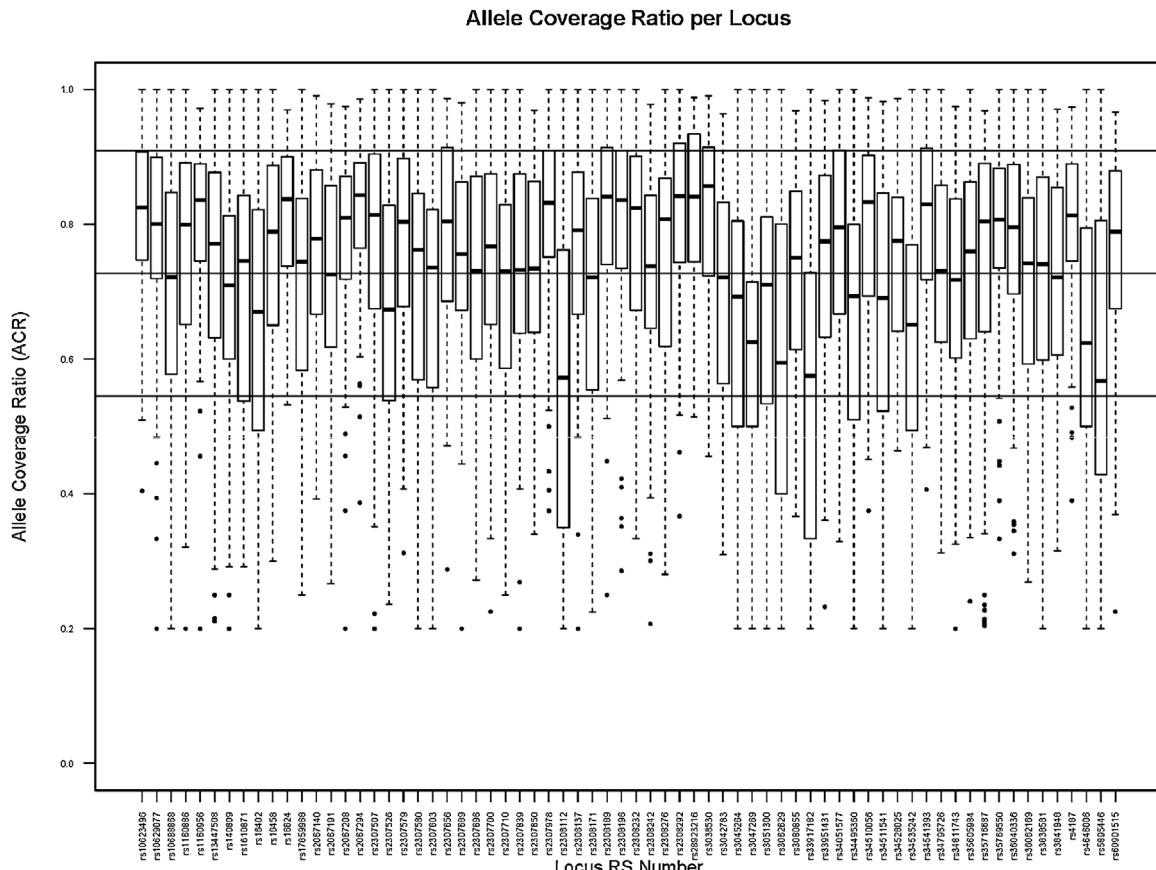


Fig. 2. Allele coverage ratio (ACR) values for 68 human identity INDELS using the Nextera™ Rapid Capture Enrichment kit and the Illumina MiSeq. Each box plot represents a single locus; the center horizontal line represents the median, the lower and upper boundaries of each box represent the first and third quartiles, respectively, the vertical dashed lines indicate plus/minus three times the interquartile range, and closed circles indicate outliers. The center horizontal line indicates the mean across 68 loci and the top and bottom horizontal lines indicate plus and minus one standard deviation, respectively for all loci combined.

Table 1

Length-based (LBAF) and sequence-based (SBAF) allele frequencies by population for 68 insertion/deletion (INDEL) markers. Target INDEL motifs are underlined and flanking region variants bolded.

INDEL RS Number	Flanking RS Number (s) and hg19 Reference Allele	Length (bp)	Sequence	AFA (N = 38)		ASA (N = 43)		CAU (N = 48)		HIS (N = 49)	
				LBAF		SBAF		LBAF		SBAF	
				0	0.3816	0.3372	0.3256	0.3438	0.3438	0.3061	0.3061
rs10623496	chr8:123945676 T ^b	100	TAAACATTGATAGCCCTTATTATATGTATGTGACACATAAACATGATTGTTCTTCTGCTTAGCAAGATTTTTCTGCTTCAG	0.3816	0.3816	0.3372	0.3256	0.3438	0.3438	0.3061	0.3061
		100	TAAACATTGATAGCCCTTATTATGTATGTGACACATAAACATGATTGTTCTTCTGCTTAGCAAGATTTTTCTGCTTCAG	0	0	0.0116	0	0	0	0	0
rs10629077	rs537464320C rs201421087C	104	TAAACATTGATAGCCCTTATTATGTATGTGACACATAAACAGAATTGATGTTCTTCTGCTTAGCAAGATTTTTCTGCTTCAG	0.6184	0.6184	0.6628	0.6628	0.6563	0.6563	0.6939	0.6939
		100	TGGTTATCTCTGCTAACTACTCTCTGCCCCATTACTACTAC <u>C</u> CTCTTCCCACAGCATCTGTCACACCTCTAAATGTTGCTCATC	0.2632	0.25	0.2791	0.2791	0.1667	0.1563	0.2755	0.2755
rs10688868 ^c	rs142634555C rs56780729C rs10902117C	100	TGGTTATCTCTGCTAACTACTCTCTGCCCCATTACTACTAC <u>C</u> CTCTTCCCACAGCATCTGTCACACCTCTAAATGTTGCTCATC	0.0132	0	0	0	0	0	0	0
		102	TGGTTATCTCTGCTAACTACTCTCTGCCCCATTACTACTAC <u>C</u> CTCTTCCCACAGCATCTGTCACACCTCTAAATGTTGCTCATC	0.7368	0.7368	0.7209	0.7209	0.8333	0.8333	0.7245	0.7245
rs1160886 ^a	–	100	CCTGTCCTCGCTAGTCCCCACTTCCATCTGCTCAGGCCCTTCATCTCACAGCCACATGGATCCACCCCTTTATGCATGTGCGAG	0.1974	0.0658	0.5698	0.4767	0.3229	0.0729	0.3673	0.1531
		102	CCTGTCCTCGCTAGTCCCCACTTCCATCTGCTCAGGCCCTTCATCTCACAGCCACATGGATCCACCCCTTTATGCATGTGCGAG	0.1316	0.093	0.25	0.25	0	0	0.2143	0.2143
rs1160956	rs138536239T	102	CCTGTCCTCGCTAGTCCCCACTTCCATCTGCTCAGGCCCTTCATCTCACAGCCACATGGATCCACCCCTTTATGCATGTGCGAG	0.8026	0.4342	0.4302	0.4186	0.6771	0.5	0.6327	0.4082
		102	CCTGTCCTCGCTAGTCCCCACTTCCATCTGCTCAGGCCCTTCATCTCACAGCCACATGGATCCACCCCTTTATGCATGTGCGAG	0.3553	0.0116	0.1667	0.1667	0	0.0104	0	0.2245
rs13447508	rs13447507A rs201219895 DEL	102	CCTGTCCTCGCTAGTCCCCACTTCCATCTGCTCAGGCCCTTCATCTCACAGCCACATGGATCCACCCCTTTATGCATGTGCGAG	0	0	0	0	0	0	0	0
		102	CAACTCTATCCCTTCCCATTGCTTAAACCTTGTGAATAGTACTCTTCTATGAAATCTGGTAGACTGACTGACTATTCTC	0.3947	0.4535	0.4063	0.3372	0	0	0	0
rs140809 ^c	rs10905513A rs687805C	100	CAACTCTATCCCTTCCCATTGCTTAAACCTTGTGAATAGTACTCTTCTATGAAATCTGGTAGACTGACTGACTATTCTC	0.6053	0.5465	0.5938	0.6628	0	0	0	0
		104	CAAATITGTCTCTCAAGGATAGCTTAAAGAGATCTCTGCTCTTAACTTCTCTGTTAGAAAAGAAAACAATACAAATTGGTTA	0.5526	0.5526	0.6047	0.6047	0.8125	0.8125	0.6125	0.6125
rs1610871 ^c	rs111817892G rs75866020C chr5:171088015 T ^b	91	CAAATITGTCTCTCAAGGATAGCTTAAAGAGATCTCTGCTCTTAACTTCTCTGTTAGAAAAGAAAACAATACAAATTGGTTA	0.4474	0.4342	0.3953	0.3953	0.1875	0.1875	0.3875	0.3875
		94	AATGTACATTATTAGATGTACTATGTTCACTGAAAGATAGCATGAACTAACAGAGTCTAATTITGACTCTCTTCTG	0.0132	0	0.0116	0	0	0	0	0
rs16402	–	100	AATGTACATTATTAGATGTACTATGTTCACTGAAAGATAGCATGAACTAACAGAGTCTAATTITGACTCTCTTCTG	0.3553	0.3553	0.5116	0.5	0.2917	0.2917	0.3854	0.3854
		100	AATGTACATTATTAGATGTACTATGTTCACTGAAAGATAGCATGAACTAACAGAGTCTAATTITGACTCTCTTCTG	0.6447	0.6184	0.4884	0.4884	0.7083	0.7083	0.6146	0.6146
rs16458	–	100	ATGTCATCTAGCATGGATTCTTCTAGGCTTAATTITACTTCAGTAAATTCAGCAGTCTGCTGTCAGTCACTCAGGAGGGGATGGCACCCAGAGTTCT	0.0263	0	0	0	0	0	0	0
		104	ATGTCATCTAGCATGGATTCTTCTAGGCTTAATTITACTTCAGTAAATTCAGCAGTCTGCTGTCAGTCACTCAGGAGGGGATGGCACCCAGAGTTCT	0.4079	0.3421	0.3605	0.186	0.4788	0.4043	0.1633	0.0816
rs16624	rs146701576C rs140263477A rs250921T	97	ATGTCATCTAGCATGGATTCTTCTAGGCTTAATTITACTTCAGTAAATTCAGCAGTCTGCTGTCAGTCACTCAGGAGGGGATGGCACCCAGAGTTCT	0.0526	0.0526	0.1628	0.0745	0	0.0612	0	0
		100	ATGTCATCTAGCATGGATTCTTCTAGGCTTAATTITACTTCAGTAAACATTCAGCAGGCTCTGTTCACTCAGGAGGGGATGGCACCCAGAGTTCT	0.0132	0.0116	0	0	0	0	0.0204	0.0204
rs17859968 ^{a,b}	rs9923304C rs16955268A	96	ATGTCATCTAGCATGGATTCTTCTAGGCTTAATTITACTTCAGTAAACATTCAGCAGGCTCTGTTCACTCAGGAGGGGATGGCACCCAGAGTTCT	0.5921	0.4474	0.6395	0.4651	0.5212	0.4574	0.8367	0.7347
		100	ATGTCATCTAGCATGGATTCTTCTAGGCTTAATTITACTTCAGTAAACATTCAGCAGGCTCTGTTCACTCAGGAGGGGATGGCACCCAGAGTTCT	0.0263	0.0263	0.1628	0.0638	0.0102	0.0102	0	0
rs2067140 ^c	rs192851878T rs254233C	100	ATTCAGAGTCATGCTGCTTAAAGAGTTGGGGATAATTACAGGCACATAAAAGCACATGGCATTAGTAGGACCCCTCAATAATGATAACTCTTACTC	0.1053	0.1053	0.0349	0	0	0.0128	0	0
		107	ATTCAGAGTCATGCTGCTTAAAGAGTTGGGGATAATTACAGGCACATAAAAGCACATGGCATTAGTAGGACCCCTCAATAATGATAACTCTTACTC	0.0132	0.0116	0	0	0	0	0	0
rs2067191	rs115923419C	100	AAAGAAAAACAGGCATGCAACTACCAACCAAAATGTCACTGACACCATGCAAAAGTCATAACATGCTGAGGAGACTGCAAGGTGAGCTATA	0.75	0.7237	0.5581	0.5581	0.2188	0.2188	0.5256	0.5612
		100	AAAGAAAAACAGGCATGCAACTACCAACCAAAATGTCACTGACACCATGCAAAAGTCATAACATGCTGAGGAGACTGCAAGGTGAGCTATA	0.0263	0	0	0	0	0	0	0
rs2067191	rs115923419C	91	GGATTAGAGTAATGTAAGTAATCTGATGAAATTACACTCTAGATAGTTATTCCTGTTGAACTACAGACATGGCATTACCCGTAACAAATTTC	0.4211	0.4211	0.4302	0.4302	0.4896	0.4896	0.3523	0.3523
		95	GGATTAGAGTAATGTAAGTAATCTGATGAAATTACACTCTAGATAGTTATTCCTGTTGAACTACAGACATGGCATTACCCGTAACAAATTTC	0.5789	0.5658	0.5698	0.5698	0.5104	0.5104	0.6477	0.6477
rs2067191	rs115923419C	95	GGATTAGAGTAATGTAAGTAATCTGATGAAATTACACTCTAGATAGTTATTCCTGTTGAACTACAGACATGGCATTACCCGTAACAAATTTC	0.0131	0	0	0	0	0	0	0

Table 1 (*Continued*)

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a Motif different from LaRue, et al.; Pereira, et al.; and qBSNP Sequences confirmed with qGV [3,6,20,22,23].

Due to lack of RS number for the observed SNP the hg19 locus coordinates are provided with hg18 coordinates, e.g., chr1:1111111-1111111, sequences contexts will be

One of twenty two INDELS with substantial sequence variation that are recommended for future HIND INDEL panels due to lack of SNPs found in the observed SNPs; the 97 locus coordinates are provided.

overall DoC mean with an average DoC of $26.5x \pm 15.7$ and overall ACR mean with an average of 0.542 ± 0.224 . While DoC and ACR values were sufficient for analysis, the rs33917182 locus was typed successfully in only 41.6% of the samples (74/178) after application of the DoC and ACR thresholds. This locus was identified by Pereira, et al. [6] as a valuable HID marker with expected heterozygosity of 0.501 and discrimination power of 0.618; LaRue, et al. [3] reported an F_{ST} of 0.0145 and observed heterozygosities ranging from 0.451 to 0.542 in four global populations for the same INDEL. Removal of this locus due to poor success resulted in full HID INDEL profiles for 155 samples, increasing the overall mean profile completion to $97.1\% \pm 0.110$.

3.2. Sequence variation

Using STRait Razor, sequence data were obtained for each INDEL motif and approximately 50 bases on either side of the motif (Table 1). Based on 1000 Genomes Project Phase 3 data [29], 100 known polymorphisms (94 SNPs and 6 non-HID INDELS) exist within 50 bases of the target HID INDELS (Supplemental Tables 3 and 4). Twenty-five and seventy-five of these polymorphisms have global allele frequencies (GAFs) ≥ 0.02 and < 0.02 , respectively. The average distance of these polymorphisms from the target INDEL was $27 \text{ bases} \pm 13$. All 25 flanking region polymorphisms with GAFs ≥ 0.02 were observed in the population data for four major US populations. Only 18/75 polymorphisms with GAFs < 0.02 were observed as would be expected due to sampling or being private variants.

In all 178 samples, 19 INDEL motifs had different sequences than previously reported, and these sequences were consistent among the samples studied herein [3,6,20]. Eighteen of these motif sequence differences were the result of alignment and were consistent with previous reports after manual analysis in IGV [22,23]. One locus, rs35716687, has been reported as a TTAA deletion but the marker was identified as a TACT deletion. Fifteen markers were associated with a repeat motif; the initial INDEL selection criteria by LaRue, et al. [3] had sought to avoid such structures by excluding loci with three or more repeats. Four of the 15 markers contained three copies or repeats. The remaining 11 loci contained two copies. These motifs range in size from di- to penta-nucleotides (Table 2). While the number of repeats is limited, STR motifs may become problematic if stutter-type artifacts can be generated. Thus, special attention during validation studies should be paid for potential stutter product generation. Though possible, STRs with only a few repeat motifs are less subject to such PCR artifacts relative to STRs with several to many repeats [30–32].

Sequence variation was observed in the region adjacent to the INDEL motif at 42 loci, producing 65 novel microhaplotypes (Table 1) [33–35]. Forty HID INDEL loci are part of a microhaplotype containing one or two SNPs. Two INDELS, rs13447508 and rs34528025, are part of microhaplotypes containing the target INDEL, an adjacent SNP (rs13447507 and rs202051643, respectively), and an adjacent flanking-region INDEL (rs201219895 and rs34247791, respectively). Twenty-two loci had sequence variants that account for $\geq 2\%$ of total alleles in two or more populations. For these 22 microhaplotypes, the presence of additional, sequence-based alleles increased the average number of alleles per marker from 2 to 3.82 ± 1.14 with a range from 3 to 7 alleles (rs1408093). The observed heterozygosity for these 22 loci increased by an average of 0.132 ± 0.0957 for AFA, 0.107 ± 0.0824 for ASA, 0.179 ± 0.106 for CAU, and 0.123 ± 0.0959 for HIS (Table 3). All 68 loci were ranked based on length- and sequence-based observed heterozygosity (Table 4). By length, INDELS rs10688868, rs2308189, rs2308276, and rs2308292 ranked 33rd, 8th, 19th, and 35nd in the HIS, AFA, ASA, and CAU populations, respectively.

Table 2

Insertion/deletion loci that are part of a short tandem repeat (STR) motif. The repeat motif for each locus is underlined.

Locus rs#	STRait Razor Sequence for Insertions	Number of Repeat Motifs	Reference
rs1160886	TAGTACTAC	2	[3,6]
rs1160956	AAAGAA <u>GAGCAAC</u>	2	[6]
rs16402	ATTAATTATTATT	2	[3,6]
rs16458	TTTACAA <u>TTCTTC</u> TT	2	[3]
rs17859968	GGCACA <u>ATAA</u> AAA	2	[3]
rs2067208	AAAGAG <u>CCCTGGCCTG</u>	2	[6]
rs2307580	TAATTAA <u>TTGAATA</u>	2	[6]
rs2307689	GGCTGT <u>TC</u> TC	3	[6]
rs2307710	CCAGAGA <u>AGGAAGGAAGGA</u>	3	[3,6]
rs2307839	TGAGAGA <u>ACAA</u> C	3	[6]
rs2307850	AGCC <u>TC</u> CCACCC	2	[3]
rs2308276	GATGA <u>TTAATT</u> AAA	2	[3]
rs3051300	AGT <u>CCATG</u> TATGT	2	[6]
rs34535242	TAGCTGGTAGGTAGGTAG	3	[3]
rs3841948	TATACAA <u>TTAATT</u> TT	2	[3]

Table 3

Length-based (LB) and sequence-based (SB) observed (H_o) and expected (H_e) heterozygosities in four major US population groups for 42 INDEL loci that exhibited sequence variation. The change in H_o and H_e as a result of utilizing SB alleles is indicated by ΔH_o and ΔH_e , respectively.

Locus	AFA						ASA					
	LB H_e	SB H_e	ΔH_e	LB H_o	SB H_o	ΔH_o	LB H_e	SB H_e	ΔH_e	LB H_o	SB H_o	ΔH_o
rs10623496	0.48	0.48	0.00	0.61	0.61	0.00	0.45	0.46	0.01	0.40	0.40	0.00
rs10629077	0.39	0.40	0.01	0.42	0.42	0.00	0.41	0.41	0.00	0.33	0.33	0.00
rs10688868	0.32	0.67	0.35	0.34	0.71	0.37	0.50	0.60	0.10	0.53	0.65	0.12
rs1160956	0.50	0.51	0.01	0.47	0.50	0.03	0.48	0.48	0.00	0.42	0.42	0.00
rs13447508	0.46	0.50	0.03	0.50	0.55	0.05	0.51	0.51	0.00	0.51	0.51	0.00
rs140809	0.49	0.68	0.19	0.34	0.45	0.11	0.47	0.70	0.24	0.40	0.51	0.12
rs1610871	0.51	0.63	0.12	0.50	0.63	0.13	0.47	0.51	0.04	0.37	0.42	0.05
rs16624	0.38	0.42	0.04	0.29	0.34	0.05	0.50	0.51	0.01	0.42	0.42	0.00
rs17859968	0.46	0.54	0.09	0.47	0.58	0.11	0.46	0.50	0.04	0.51	0.53	0.02
rs2067140	0.35	0.65	0.29	0.39	0.66	0.26	0.46	0.52	0.06	0.42	0.49	0.07
rs2067191	0.49	0.51	0.02	0.53	0.55	0.03	0.50	0.50	0.00	0.58	0.58	0.00
rs2067208	0.32	0.32	0.00	0.34	0.34	0.00	0.29	0.35	0.06	0.26	0.28	0.02
rs2307507	0.32	0.33	0.00	0.29	0.29	0.00	0.45	0.45	0.00	0.41	0.41	0.00
rs2307526	0.47	0.67	0.20	0.53	0.71	0.18	0.47	0.50	0.03	0.54	0.54	0.00
rs2307579	0.49	0.50	0.01	0.50	0.50	0.00	0.29	0.29	0.00	0.30	0.30	0.00
rs2307656	0.48	0.55	0.07	0.45	0.50	0.05	0.50	0.58	0.08	0.47	0.56	0.09
rs2307689	0.50	0.59	0.09	0.58	0.61	0.03	0.40	0.66	0.27	0.40	0.72	0.33
rs2307700	0.46	0.67	0.21	0.42	0.61	0.18	0.40	0.48	0.08	0.35	0.42	0.07
rs2307978	0.48	0.67	0.18	0.47	0.68	0.21	0.49	0.66	0.18	0.44	0.60	0.16
rs2308189	0.50	0.76	0.25	0.55	0.82	0.26	0.50	0.64	0.14	0.60	0.74	0.14
rs2308232	0.38	0.39	0.01	0.39	0.39	0.00	0.40	0.43	0.03	0.39	0.44	0.05
rs2308242	0.45	0.45	0.00	0.29	0.29	0.00	0.40	0.40	0.00	0.49	0.49	0.00
rs2308276	0.50	0.53	0.03	0.53	0.55	0.03	0.49	0.65	0.16	0.49	0.77	0.28
rs2308292	0.50	0.59	0.08	0.50	0.61	0.11	0.50	0.64	0.14	0.47	0.60	0.14
rs28923216	0.44	0.45	0.02	0.37	0.39	0.03	0.50	0.50	0.00	0.50	0.50	0.00
rs3038530	0.46	0.59	0.13	0.47	0.63	0.16	0.47	0.65	0.19	0.58	0.67	0.09
rs3042783	0.35	0.50	0.14	0.34	0.50	0.16	0.48	0.53	0.05	0.53	0.56	0.02
rs3045264	0.41	0.41	0.00	0.50	0.50	0.00	0.46	0.46	0.00	0.43	0.43	0.00
rs3051300	0.30	0.30	0.00	0.26	0.26	0.00	0.48	0.48	0.00	0.44	0.44	0.00
rs33951431	0.44	0.68	0.24	0.37	0.61	0.24	0.48	0.55	0.07	0.49	0.58	0.09
rs34511541	0.49	0.60	0.11	0.34	0.42	0.08	0.51	0.59	0.08	0.58	0.65	0.07
rs34528025	0.50	0.50	0.00	0.42	0.42	0.00	0.50	0.50	0.00	0.51	0.51	0.00
rs34795726	0.49	0.50	0.02	0.29	0.29	0.00	0.43	0.43	0.00	0.37	0.37	0.00
rs34811743	0.49	0.56	0.07	0.55	0.63	0.08	0.31	0.31	0.00	0.14	0.14	0.00
rs35605984	0.50	0.50	0.00	0.63	0.63	0.00	0.49	0.49	0.00	0.45	0.45	0.00
rs35769550	0.19	0.21	0.02	0.21	0.24	0.03	0.50	0.50	0.00	0.49	0.49	0.00
rs36062169	0.51	0.56	0.06	0.66	0.68	0.03	0.43	0.44	0.02	0.47	0.49	0.02
rs3838581	0.43	0.44	0.01	0.39	0.39	0.00	0.45	0.45	0.00	0.49	0.49	0.00
rs3841948	0.48	0.57	0.08	0.32	0.45	0.13	0.51	0.51	0.00	0.60	0.60	0.00
rs4646006	0.29	0.29	0.00	0.29	0.29	0.00	0.49	0.50	0.01	0.44	0.47	0.02
rs5895446	0.45	0.56	0.11	0.50	0.61	0.11	0.44	0.65	0.21	0.42	0.60	0.19
rs60901515	0.46	0.51	0.05	0.38	0.43	0.05	0.44	0.47	0.03	0.44	0.47	0.02
Locus	CAU						HIS					
rs10623496	LB H_e	SB H_e	ΔH_e	LB H_o	SB H_o	ΔH_o	LB H_e	SB H_e	ΔH_e	LB H_o	SB H_o	ΔH_o
rs10629077	0.46	0.46	0.00	0.31	0.31	0.00	0.43	0.43	0.00	0.41	0.41	0.00
rs10688868	0.28	0.28	0.00	0.29	0.29	0.00	0.40	0.40	0.00	0.39	0.39	0.00
rs1160956	0.44	0.66	0.22	0.44	0.65	0.21	0.47	0.72	0.25	0.45	0.76	0.31
rs13447508	0.42	0.42	0.00	0.33	0.33	0.00	0.48	0.48	0.00	0.48	0.48	0.00
rs140809	0.50	0.62	0.12	0.45	0.55	0.11	0.28	0.45	0.17	0.24	0.41	0.16
rs1610871	0.49	0.49	0.00	0.54	0.54	0.00	0.51	0.52	0.01	0.38	0.41	0.03

Table 3 (Continued)

Locus	AFA						ASA					
	LB H _e	SB H _e	ΔH _e	LB H _o	SB H _o	ΔH _o	LB H _e	SB H _e	ΔH _e	LB H _o	SB H _o	ΔH _o
rs16624	0.35	0.36	0.02	0.27	0.29	0.02	0.50	0.51	0.01	0.59	0.59	0.00
rs17859968	0.50	0.50	0.00	0.54	0.54	0.00	0.49	0.52	0.02	0.51	0.55	0.04
rs2067140	0.48	0.55	0.07	0.52	0.56	0.04	0.47	0.55	0.07	0.47	0.53	0.06
rs2067191	0.51	0.51	0.00	0.40	0.40	0.00	0.46	0.46	0.00	0.43	0.43	0.00
rs2067208	0.43	0.59	0.16	0.38	0.54	0.17	0.43	0.50	0.07	0.45	0.51	0.06
rs2307507	0.49	0.49	0.00	0.48	0.48	0.00	0.50	0.50	0.00	0.37	0.37	0.00
rs2307526	0.50	0.61	0.10	0.52	0.63	0.10	0.42	0.63	0.21	0.45	0.73	0.27
rs2307579	0.49	0.49	0.00	0.44	0.44	0.00	0.46	0.46	0.00	0.35	0.35	0.00
rs2307656	0.51	0.61	0.10	0.63	0.71	0.08	0.46	0.52	0.06	0.48	0.50	0.02
rs2307689	0.35	0.63	0.28	0.40	0.69	0.29	0.47	0.67	0.21	0.44	0.56	0.12
rs2307700	0.50	0.61	0.11	0.50	0.63	0.13	0.43	0.59	0.15	0.38	0.52	0.15
rs2307978	0.28	0.62	0.34	0.25	0.67	0.42	0.46	0.66	0.20	0.29	0.53	0.24
rs2308189	0.46	0.67	0.21	0.40	0.70	0.30	0.50	0.61	0.10	0.57	0.62	0.05
rs2308232	0.42	0.46	0.04	0.51	0.53	0.02	0.42	0.45	0.04	0.37	0.42	0.05
rs2308242	0.34	0.36	0.02	0.43	0.43	0.00	0.35	0.35	0.00	0.35	0.35	0.00
rs2308276	0.50	0.50	0.00	0.54	0.54	0.00	0.50	0.54	0.04	0.54	0.59	0.05
rs2308292	0.42	0.70	0.28	0.46	0.75	0.29	0.44	0.68	0.24	0.47	0.69	0.22
rs28923216	0.50	0.50	0.00	0.48	0.48	0.00	0.50	0.50	0.00	0.53	0.53	0.00
rs3038530	0.46	0.67	0.21	0.35	0.48	0.13	0.50	0.65	0.14	0.51	0.64	0.13
rs3042783	0.40	0.55	0.15	0.42	0.60	0.19	0.50	0.58	0.08	0.57	0.63	0.06
rs3045264	0.46	0.47	0.01	0.48	0.48	0.00	0.45	0.45	0.00	0.56	0.56	0.00
rs3051300	0.50	0.50	0.00	0.51	0.51	0.00	0.47	0.48	0.01	0.43	0.45	0.02
rs33951431	0.49	0.49	0.00	0.52	0.52	0.00	0.47	0.52	0.05	0.47	0.51	0.04
rs34511541	0.44	0.66	0.22	0.35	0.54	0.19	0.50	0.60	0.09	0.57	0.61	0.04
rs34528025	0.45	0.45	0.00	0.46	0.46	0.00	0.44	0.56	0.12	0.39	0.49	0.10
rs34795726	0.50	0.51	0.01	0.50	0.52	0.02	0.48	0.48	0.00	0.46	0.46	0.00
rs34811743	0.45	0.45	0.00	0.33	0.33	0.00	0.37	0.41	0.05	0.40	0.46	0.06
rs35605984	0.50	0.51	0.01	0.46	0.48	0.02	0.49	0.49	0.00	0.51	0.51	0.00
rs35769550	0.50	0.50	0.00	0.60	0.60	0.00	0.47	0.47	0.00	0.41	0.41	0.00
rs36062169	0.49	0.49	0.00	0.50	0.50	0.00	0.50	0.50	0.00	0.59	0.59	0.00
rs3838581	0.44	0.44	0.00	0.44	0.44	0.00	0.51	0.51	0.00	0.47	0.47	0.00
rs3841948	0.47	0.47	0.00	0.54	0.54	0.00	0.42	0.42	0.00	0.38	0.38	0.00
rs4646006	0.49	0.49	0.00	0.55	0.55	0.00	0.50	0.50	0.00	0.39	0.39	0.00
rs5895446	0.47	0.66	0.19	0.52	0.73	0.21	0.44	0.67	0.23	0.24	0.55	0.31
rs60901515	0.45	0.45	0.00	0.58	0.58	0.00	0.49	0.50	0.01	0.61	0.61	0.00

However, when ranked by sequence-based observed heterozygosity, microhaplotypes containing these four INDELS displayed the highest heterozygosities in the HIS, AFA, ASA, and CAU populations, respectively. The second highest heterozygosity microhaplotypes in the AFA, HIS, ASA, and CAU populations are rs10688868, rs2307526, rs2308189, and rs5895446, respectively. These four loci increased from their length-based ranks of 52nd, 32nd, 3rd, and 16th, in AFA, HIS, ASA, and CAU, respectively. Microhaplotypes containing the rs10688868 and rs2308189 INDELS are ranked highest, or second highest, in heterozygosity in the AFA, ASA, and HIS populations, making them far more informative than even the top ranked length-based marker. Single-locus RMPs were decreased by an average of 0.166 ± 0.0816 for AFA, 0.130 ± 0.0661 for ASA, 0.176 ± 0.0837 for CAU, and 0.134 ± 0.0773 for HIS (Supplemental Table 5).

The remaining 20 loci with detectable adjacent sequence variants did not display substantial sequence variation (average frequency of 0.0234 ± 0.0250 across all four populations). It should be noted that there were specific microhaplotypes containing INDELS rs34528025, rs36062169, and rs3841948 with relatively high frequencies: 0.1020 for HIS, 0.0658 for AFA, and 0.07900 for ASA, respectively. However, they were either observed once or not at all in the other population groups. While microhaplotypes containing these three INDELS did not substantially increase the discrimination power across the populations, these alleles may hold value for ancestry apportionment. The low allele frequency of these sequence variants, or lack of sufficient frequency in multiple populations, suggests that these 20 microhaplotypes do not have increased discrimination power over that of the current length-based allele polymorphism (Table 1) for HID applications.

Length-based allele frequencies and observed and expected heterozygosities were similar to those previously reported by LaRue, et al. [3] and Pereira, et al. [6]. Prior to Bonferroni correction, three AFA, three ASA, no CAU, and three HIS length-based loci and four AFA, five ASA, three CAU, and two HIS sequence-based loci deviated significantly from HWE ($p < 0.05$). After Bonferroni correction, there were no significant departures from HWE for length- or sequence-based loci ($p = 0.00074$, Supplemental Table 6). Prior to Bonferroni correction, 185 AFA, 140 ASA, 197 CAU, and 216 HIS length-based and 205 AFA, 186 ASA, 124 CAU, and 186 HIS sequence-based pairwise LDs were observed ($p = 0.05$). Five (AFA), four (ASA), seven (CAU), and five (HIS) length-based and seven (AFA), eight (ASA), nine (CAU), and six (HIS) sequence-based significant pairwise LDs were observed for markers on the same chromosome but not on the same chromosomal arm. After Bonferroni correction, at most two pairwise locus comparisons showed significant LD for length- and sequence-based alleles per population (rs2308112 and rs34795726 in AFA, rs34541393 and rs34811743 in ASA, $p < 0.0000219$). The observed significant pairwise LDs are less than that due to chance alone (~114). Assuming independence, the combined length-based RMPs were 1.36×10^{-26} for AFA, 5.42×10^{-27} for ASA, 2.94×10^{-27} for CAU, 1.33×10^{-27} for HIS and the combined sequence-based RMPs were 3.29×10^{-32} for AFA, 5.92×10^{-31} for ASA, 6.69×10^{-32} for CAU, and 5.67×10^{-32} for HIS for 68 HID INDEL-containing microhaplotypes (Supplemental Table 5).

The combined RMPs, under the assumption of independence, for 22 microhaplotypes (Supplemental Table 5) were 3.84×10^{-14} for AFA, 3.87×10^{-13} for ASA, 7.76×10^{-14} for CAU, and 1.60×10^{-13} for HIS. These values are comparable to those obtained with larger INDEL panels described by Pereira, et al. [6] and LaRue, et al. [3].

Table 4

Length-based (LB) and sequence-based (SB) observed heterozygosity rank (1=highest) in four major US population groups for 68 INDEL loci.

Locus	AFA		ASA		CAU		HIS	
	LB Rank	SB Rank						
rs10623496 ^a	5	13	50	55	64	65	44	51
rs10629077 ^a	38	46	59	59	65	66	47	54
rs10688868 ^{a,b}	52	2	9	6	45	7	33	1
rs1160886	10	24	10	18	28	39	37	44
rs1160956 ^a	24	26	42	47	63	64	23	34
rs13447508 ^a	15	21	12	20	62	62	21	33
rs140809 ^{a,b}	53	41	51	21	41	18	67	52
rs1610871 ^{a,b}	16	7	54	48	10	20	50	49
rs16402	11	25	65	65	61	63	58	61
rs16458	25	34	43	49	36	45	24	35
rs16624 ^a	58	57	44	50	66	67	5	12
rs17859968 ^{a,b}	26	19	13	19	11	21	16	19
rs2067140 ^{a,b}	42	6	45	25	17	16	26	21
rs2067191 ^a	12	22	4	12	52	56	40	46
rs2067208 ^{a,b}	54	58	66	66	56	27	34	25
rs2067294	64	64	46	51	40	49	20	32
rs2307507 ^a	59	59	49	54	33	44	57	60
rs2307526 ^{a,b}	13	3	8	17	15	9	32	2
rs2307579 ^a	17	27	62	62	42	50	59	62
rs2307580	65	65	23	31	25	34	39	45
rs2307603	27	35	60	60	3	12	10	15
rs2307656 ^{a,b}	35	28	24	14	2	3	22	28
rs2307689 ^{a,b}	6	14	52	3	51	5	38	17
rs2307696	39	47	25	32	55	59	2	7
rs2307700 ^{a,b}	40	15	57	52	23	10	54	24
rs2307710	2	8	30	37	5	14	56	59
rs2307839	63	63	17	26	67	68	61	64
rs2307850	28	36	31	38	43	51	53	58
rs2307978 ^{a,b}	29	4	32	8	68	6	64	23
rs2308112	66	66	1	4	19	31	51	56
rs2308137	43	50	33	39	31	42	62	65
rs2308171	48	55	67	67	57	60	65	67
rs2308189 ^{a,b}	8	1	3	2	49	4	9	8
rs2308196	30	37	47	53	37	46	35	41
rs2308232 ^{a,b}	44	51	53	44	20	28	55	48
rs2308242 ^a	60	60	18	27	46	53	60	63
rs2308276 ^{a,b}	14	23	19	1	9	22	13	13
rs2308292 ^{a,b}	18	16	26	9	35	1	27	3
rs28923216 ^a	49	52	16	24	32	43	14	22
rs3038530 ^{a,b}	31	9	5	5	58	37	15	4
rs3042783 ^{a,b}	55	29	11	15	48	11	7	5
rs3045264 ^a	19	30	40	45	29	40	11	16
rs3047269	7	20	41	46	26	35	4	11
rs3051300 ^a	67	67	34	40	21	32	42	43
rs3062629	32	38	27	33	27	36	18	29
rs3080855	36	42	14	22	12	23	28	37
rs33917182	50	56	58	58	50	55	66	68
rs33951431 ^{a,b}	51	17	20	13	18	29	29	26
rs34051577	3	10	35	41	47	54	30	38
rs34495360	46	54	7	16	54	58	63	66
rs34510056	20	31	63	63	1	8	48	55
rs34511541 ^{a,b}	56	48	6	7	59	26	8	9
rs34528025 ^a	41	49	15	23	39	48	49	30
rs34535242	33	39	55	56	30	41	12	18
rs34541393	37	43	61	61	13	24	36	42
rs34795726 ^a	61	61	56	57	24	30	31	39
rs34811743 ^{a,b}	9	11	68	68	60	61	45	40
rs35605984 ^a	4	12	29	36	34	38	17	27
rs35716687	21	32	36	42	7	17	41	47
rs35769550 ^a	68	68	21	28	4	13	43	50
rs36040336	22	33	64	64	53	57	19	31
rs36062169 ^a	1	5	28	29	22	33	6	14
rs3838581 ^a	45	53	22	30	44	52	25	36
rs3841948 ^a	57	44	2	10	14	25	52	57
rs4187	34	40	37	43	38	47	1	6
rs4646006 ^a	62	62	38	34	8	19	46	53
rs5895446 ^{a,b}	23	18	48	11	16	2	68	20
rs60901515a ^b	47	45	39	35	6	15	3	10

^a Marker is part of a microhaplotype observed in these population data.

^b Marker is recommended for future massively parallel sequencing HID microhaplotype panels.

4. Conclusion

Sixty-eight HID INDELS were characterized further using MPS and a novel application of the STRait Razor software. Fifteen loci were found to be part of an STR, and as such, although unlikely, special attention to potential stutter artifacts should be given with these markers for PCR-based sample preparation on MPS platforms.

The presence of additional, sequence-based alleles in 42 microhaplotypes increased heterozygosities and decreased the single-locus random match probabilities in four major U.S. populations. A subset of 22 sequence-based microhaplotype alleles became substantially more informative for identity testing than solely their length-based equivalents. The remaining 20 loci had less frequent sequence variation: variants were observed at a frequency <0.02 in one or more populations (10 loci) or at a frequency ≥0.02 in one major US population (7 loci). While not increasing discrimination power substantially, the relatively common alleles seen in only one population group may be informative for ancestry determination.

The sample population sizes used for this exploratory study are less than those typically used for STR population studies. However, there are far less alleles per locus for INDELS and INDELS within microhaplotypes. As described previously by Chakraborty [36] economization of sample size for allele characterization can be achieved by focusing on obtaining reliable frequencies of common alleles in a sample, and rare allele frequencies can be approximated by an upper bound. Since microhaplotype alleles with reasonable minimum allele frequencies of, for example $p=0.05$ with an $\alpha=0.05$, can be detected in the sample sizes in this study, the population data are reasonable for identifying markers that likely provide increased discrimination power over using just the INDEL itself.

This study focused on markers that could be converted to short amplicons which would be more beneficial for degraded DNA sample typing than would be STRs. The positions of all INDELS are already known and readily accessible. Therefore, STR population data to test for linkage disequilibrium were not considered in the current study. In addition, to perform such a study a larger population sample would be required to accommodate the more polymorphic STRs.

Approximately 50 bases on either side of the INDEL were searched so that microhaplotype amplicon sizes could be designed to be as short as possible. In this dataset, multiple polymorphisms were captured on the same amplicon so future studies may focus on phasing assessments to aid in mixture deconvolution. While a relatively short amplicon is desirable for analyzing challenged samples, it is possible that additional polymorphisms lie beyond the flanking regions analyzed herein.

The Nextera™ Rapid Capture was used to readily identify markers and variants without the demands of primer design associated with PCR-amplicon enrichment. In addition, while 50 ng of genomic DNA is perfectly acceptable to use for exploratory work, this amount of input DNA clearly is far too much for an assay for forensic utility. It is expected that those microhaplotypes of interest will be converted to an assay that is PCR-based and thus requires input DNA of ≤ 1 ng.

CE and MPS platforms are suitable for analysis of INDELS; however, with MPS, 42 markers had increased variation due to closely linked polymorphisms. The panel of 22 INDEL-containing microhaplotypes had increased numbers of alleles, combined RMPs comparable to those provided by larger INDEL sets in LaRue, et al. (38 and 49 INDELS) [3] and Pereira, et al. (38 INDELS) [6], and heterozygosities greater than some low-performing STR markers [37,38].

Conflict of interest

The authors report no conflict of interest.

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Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.fsigen.2016.09.005>.

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